



**Hong Kong Institute of Medical Laboratory Sciences**

**Quality Assurance Programme**

***February Survey (2026)***

Anatomical Pathology

# HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES

## QUALITY ASSURANCE PROGRAMME

### Anatomical Pathology

#### INSTRUCTIONS

1. All tests should be performed as soon as possible after the receipt of the specimens.
2. Unless otherwise specified, all survey material had been fixed in 10% buffered formalin and paraffin embedded.
3. The survey material should be treated, as far as possible, like a patient specimen.
4. The survey material should be treated as potentially hazardous and standard laboratory safety precautions should be taken as usual in your laboratory.
5. To complete the survey, you should send back the stained slides together with appropriate controls, and the questionnaire for statistical analysis and compilation of results.
6. Do NOT forward any particulars of your laboratory except your assigned confidential Laboratory Code. **Late returns will be counted as not done and the results will NOT be accepted for assessment and statistical analysis.**
7. All stained slides should be properly labelled with the **assigned 4-digit laboratory code but not the identity of your laboratory.**
8. Place the labelled slides inside the slide mailers.
9. Return the slide mailers and complete questionnaire in the enclosed foam-padded envelope.
10. Arrange return of survey material with the courier service provider according to the instruction as per attached before on **26 March 2026**

For further enquiry, please contact HKIMLSQAP Ltd.

Phone: (852) 2499 0015 Fax: (852) 2124 2798 E-mail: [info@hkimlsqap.org](mailto:info@hkimlsqap.org)

#### CONFIDENTIALITY

HKIMLSQAP is committed to keeping all details of participants confidential. Please refer to <http://www.hkimlsqap.org>

**HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES  
QUALITY ASSURANCE PROGRAMME  
ANATOMICAL PATHOLOGY**

**FEBRUARY SURVEY (2026)**

**HISTOLOGICAL STAINING MODULE**

You are provided with two paraffin sections, which had been fixed in 10% buffered formalin. Perform the following staining methods and return the **STAINED SECTIONS** for assessment.

**HC2601** Stain with your routine H&E method

**HC2602** Stain with Martius, Scarlet and Blue (MSB) method to demonstrate fibrin

**POSITIVE CONTROLS ARE REQUIRED BUT NOT FOR ASSESSMENT.**

Please label the slides with the corresponding **Test code** and your confidential **Laboratory Code** for identification purposes. **No mark will be given for the unlabelled slide.** Put not more than two slides in one mailer in each return. Please ensure the mountant is completed dried before putting into the mailer.



**HISTOLOGICAL STAINING MODULE  
RETURN FORM**

Laboratory Code:

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Date of Return: on or before **26 March 2026**

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**QUESTIONNAIRE**

**SECTION I: HISTOPATHOLOGY**

1. What is the section thickness for your routine fibrin stain?

- 3 µm \_\_\_\_\_
  - 4 µm \_\_\_\_\_
  - 5 µm \_\_\_\_\_
  - 6 µm \_\_\_\_\_
  - Others (please state) \_\_\_\_\_
- 

2. Do you use MSB method for demonstrating fibrin?

- Yes \_\_\_\_\_
  - No \_\_\_\_\_
  - If NO, please specify the method \_\_\_\_\_
- 

3. Do you use commercial MSB solution for demonstrating fibrin?

- Yes \_\_\_\_\_
  - No \_\_\_\_\_
  - If Yes, please specify the manufacturer \_\_\_\_\_
- 

4. Do you have a Mordant step in routine fibrin staining?

- Yes \_\_\_\_\_
- No \_\_\_\_\_

If Yes, please state the concentration of Mordant solution used and the staining time:

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Laboratory Code:

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Date of Return: on or before **26 March 2026**

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5. How long will you keep the staining dye of Martius yellow, Brilliant Crystal Scarlet and Blue solution?

Freshly Prepared \_\_\_\_\_

1 day \_\_\_\_\_

1 week \_\_\_\_\_

1 month \_\_\_\_\_

Others (please state) \_\_\_\_\_

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6. Do you stain the nuclei with Celestine blue – haematoxylin solution?

Yes \_\_\_\_\_

No \_\_\_\_\_

Others (please state) \_\_\_\_\_

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7. Do you filter the working staining dyes before use?

Yes \_\_\_\_\_

No \_\_\_\_\_

Others (please state) \_\_\_\_\_

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8. Do you rise in 95% alcohol before stain in Martius yellow?

Yes \_\_\_\_\_

No \_\_\_\_\_

Others (please state) \_\_\_\_\_

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9. How long will you be treated with phosphotungstic acid (PTA)?

1 – 5 minutes \_\_\_\_\_

5 – 10 minutes \_\_\_\_\_

10 – 30 minutes \_\_\_\_\_

>30 minutes \_\_\_\_\_

Others (please state) \_\_\_\_\_

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10. Methyl blue or soluble blue solution used in your laboratory after PTA staining?

Methyl blue \_\_\_\_\_

Soluble blue \_\_\_\_\_

Others (please state) \_\_\_\_\_

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Laboratory Code: 

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Date of Return: on or before **26 March 2026**

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11. Do you rinse in acetic acid after **Methyl blue/Soluble blue/Others** staining?

Yes \_\_\_\_\_  
 No \_\_\_\_\_  
 Others (please state) \_\_\_\_\_

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12. Do you blot dry the slide before dehydrating, clear and mount?

Yes \_\_\_\_\_  
 No \_\_\_\_\_  
 Others (please state) \_\_\_\_\_

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**MSB Method**

<i>Reagent</i>	<i>% / Composition</i>	<i>Time</i>	<i>Temp (°C)</i>
Mordant (If Yes)	_____ % _____ _____ % _____	min.	
Martius yellow Solution	Martius yellow _____ g Phosphotungstic acid _____ g _____ % Alcohol _____ ml	min.	
Brilliant Crystal Scarlet Solution	Brilliant Crystal Scarlet _____ g _____ ml _____ ml	min.	
Phosphotungstic acid (PTA) Solution	Phosphotungstic acid _____ g Water _____ ml	min.	
_____ Blue Solution	_____ Blue _____ g _____ ml _____ ml	min.	

**END**

HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES  
QUALITY ASSURANCE PROGRAMME  
ANATOMICAL PATHOLOGY

FEBRUARY SURVEY (2026)

**IMMUNOHISTOCHEMICAL STAINING MODULE**

**Evaluation of CK20 Antibody**

**HC2603** You are provided with a paraffin section, stain the slide with your **in-house Anti-Human CK20** antibody and your routine immunohistochemistry detection system.

**Evaluation of CDX2 Demonstration**

**HC2604** You are provided with a paraffin section, stain the slide with your **in-house Anti-Human CDX2** antibody and your routine immunohistochemistry detection system.

**Evaluation of AE1/3 Demonstration**

**HC2605** You are provided with a paraffin section, stain the slide with your **in-house Anti-Human AE1/3** antibody and your routine immunohistochemistry detection system.

**POSITIVE CONTROLS ARE REQUIRED BUT NOT FOR ASSESSMENT.**

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IMMUNOHISTOCHEMICAL STAINING MODULE  
 RETURN FORM

Laboratory Code: 

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Date of Return: on or before **26 March 2026**

**QUESTIONNAIRE**

Your immunohistochemistry conditions:

<i>STEP</i>	<i>HC2603 (CK20 in-house)</i>	<i>HC2604 (CDX2 in-house)</i>	<i>HC2605 (AE1/3 in-house)</i>
Supplier			
Dilution	1:	1:	1:
Peroxidase Blocking	min.	min.	min.
Antigen retrieval: <input type="checkbox"/> Trypsinization <input type="checkbox"/> Microwave <input type="checkbox"/> Pressure cooking <input type="checkbox"/> PT module <input type="checkbox"/> Others	YES / NO  min.	YES / NO  min.	YES / NO  min.
Detection System			
Duration of Colour Development	DAB min.	DAB min.	DAB min.
End product Colour Enhancement (if any)	min.	min.	min.

END